# **Horticulture Lighting with LEDs**



OS SSL | November 2016 | Regensburg **Light is OSRAM**

# **Horticulture Lighting What is horticulture lighting and how is it used?**

- **Supplemental Lighting** To supplement natural daylight and raise grow light levels in order to enhance photosynthesis and thereby improve growth and quality of plants in greenhouses.
- **Photoperiodic Lighting**

To control the light period by extending the natural day length with artificial light.

• **Cultivation without daylight** To totally replace daylight with artificial light for ultimate climate control.







#### **Horticulture Lighting What is horticulture lighting and how is it used?**

**Horticulture lighting is used to support, increase and enable the growth of plants by illuminating them with artificial light. LED Light is a very efficient way and upcoming solution for this application!**





#### **Top Lighting Inter Lighting Vertical Farming**





## **OSLON® SSL Green House Lighting with LEDs**

#### **Toplighting**



Toplighting is currently used with conventional light sources. The plants are illuminated from the top similar sun light.

The high power consumption and the heat of HPS luminaires are also demanding a distance between the light source and the plants.

#### **Interlighting**



Interlighting is enabled by LEDs as a light source! In this case the lighting is in between the plants and leaves. This should reduce the shadowing of the leafs which may occur by top lighting. This increases the amount of light even on the lower leafs.

Unlike the hot HPS Luminaires, the comparatively low temperatures on the LED luminaire don't damage the plants.



# **Horticulture Lighting How does light affect the plant growth?**

#### • **Light quantity**

The amount of light affects the photosynthesis process in the plant. This process is a photochemical reaction within the chloroplasts of the plant cells in which CO2 is converted into carbohydrate under the influence of the light energy.

#### • **Light quality regarding spectral composition of the light**

The spectral composition of the different wavelength regions (blue, green, yellow, red, far red or invisible e.g. UV or IR) is important for the grows, shape, development and flowering (photomorphogenesis) of the plant. For the photosynthesis, the blue and red regions are most important.

#### • **Light duration**

The timing / light duration which is also called photoperiod is mainly affecting the flowering of the plants. The flowering time can be influenced by controlling the photoperiod.

**Source: [0];[18]**



## **Difference in absorption curves for photochemical reactions between the human eye and plants**

**Light is generating a photochemical reaction. In our eye it is reacting with the photo receptor in different versions S, M and L. In plants, the light is reacting with Chlorophyll a and b.**





#### **Different receptors – different units**



**Horticulture Lighting with LEDs | OS SSL | NR AW CH** OSRAM Opto Semiconductors| November 2016 7



### **Photosynthetic efficiency is mainly driven by chlorophyll a and b**

• **Chlorophyll a and b** Mainly responsible for the photosynthesis and responsible for the definition of the area for the photosynthetically active radiation PAR.

#### •**Carotenoid**

Further photosynthetic pigments also known as antenna pigments like carotenoids  $\beta$ -carotene, zeaxanthin, lycopene and lutein etc.



#### **Source: [18],[19]**



#### **Horticulture Lighting 450nm and 660nm provide the energy for the plant**

**The 450nm and the 660nm are providing the energy for the plant to life and grow. The amount of light is not measured in lumen but in amount of photons. The common unit in horticulture lighting is µmol/s in the range of 400-700 (photosynthetically active region)**



**PAR 400 – 700nm**

Usually the customer will request for a certain photon flux level in µmol/s. The values can be put in our horticulture calculator to derive the number of LEDs

[Horticulture System Calculator](https://lightweb.osram-light.com/content/10003469/Applications/Application_Knowledge/Blockbuster/Horticulture Lighting/160415 Horticultural System Calculator.xlsx?Web=1)



## **Photomorphogenic effects are mainly influenced by the phytochromes Pr and Pfr**

• **Phytochrome Pr and Pfr**

The Phytochromes pr (red) and pfr (far red) are mainly influencing the germination, plant growth, leave building and flowering.

#### •**Phytomorphogenic effects**

The phytomorphogenic effects are controlled by applying a spectrum with a certain mix of 660nm and 730nm in order to stimulate the pr and pfr phytochromes.





## **Horticulture Lighting - The 730nm is necessary to control the growth of the plant**

**The application of the 730nm is much more complicated and needs special knowledge by the grower. The 730nm LED should be in a separate string and dimmable in the luminaire.**



The 730nm LEDs can be used to influence the length growth of the plant. Shining 730nm light on a plant makes the plant feel like being in the shadow of a bigger plant and triggers the "shade escape reaction" which means it grows very fast.

Another effect which can be influenced by the 730nm LEDs is the timing of blossoming of flowers. It can make the flowers blossom in winter time of even prevent the blossoming in summer time.



# **Special potential of LEDs in floriculture lighting**

**Traditionally ornamental plants are of high economic importance. The Red and Far-Red light mediates the conversion of phytochromes which can control the triggers for flowering.**

**Illumination with 730nm:** The cycle from Pr to Pfr is initiated by red light of 660nm which represents daylight. During the night time, the Pfr is converted back to Pr. This back conversion can also be actively be influenced by 730nm far red light.

This enables a perfect control of the flowering timing independent of the seasons.





## **Therefore we are focusing in horticulture lighting on the 450nm, 660nm and 730nm LEDs**

**All three important wavelength are available in the same LED package:**



**Horticulture Lighting with LEDs | OS SSL | NR AW CH** OSRAM Opto Semiconductors| November 2016 13



## **The incumbent – High-Pressure Sodium (HPS) lamps**

**Today's widely used High-Pressure Sodium lamps produce over 100 lm/W, but over a wide wavelength range**

**Efficacy in Lumen per Watt is misleading, since plants don't have eyes**

**Typical lifetime is (only) ~8000h**

**Takes minutes to reach full power**

**Large lamps are most cost efficient**





### **Horticulture Lighting - What are the LEDs and colors used for horticulture lighting?**

**The typical wavelength used for horticulture lighting are 450nm and 660nm. For the control of the plants 730nm are used**





#### **One spectrum and three different definitions of the wavelength**

1,00





Watts/nm

## **Background Knowledge Photon counting**

**Today's method of weighing the spectrum is not really adequate** 





## **Background Knowledge Photon counting**

**Today's method of weighing the spectrum is not really adequate** 





# **Effect of the different wavelength regions on plants**

**Different regions of the wavelength in the illumination spectrum have different effects on the plants:**





## **What are typical µmol/s.m² values for horticulture lighting?**







#### **What light level for what cut flower?**



#### **Source: http://www.hortilux.nl/light-technology**



# **Effect of red light around 660nm on physiology of vegetables**





# **Effect of red light around 660nm on physiology of vegetables**





## **Effect of red light around 660nm on physiology of vegetables**





## **Effect of blue light around 450nm on physiology of vegetables**



## **Effect of green light around 520nm on physiology of vegetables**



## **Horticulture Lighting Example LED light ratios for different purposes**



The highest efficacy of  $\mu$ mol/J from the spectrum can be achieved by using the 660nm Red LEDs combined with some 450nm Blue LEDs to maintain a reasonable ratio between the wavelengths



Especially for growth of the leafy green vegetable plants the vegetative growth ratio is used to achieve fastest growth where visible assessment of plant health is not important





**Source: http://www.illumitex.com/illumitex-leds/surexi-horticulture-leds/**



### **Horticulture Lighting Example LED light ratios for different purposes**



A high blue content in the spectrum is recommended for growth of the seedlings.



**Source: http://www.illumitex.com/illumitex-leds/surexi-horticulture-leds/**



## **Horticulture Lighting Example LED light ratios for different purposes**

#### **Actual Studies:**

• **More kind of wavelegth may be needed by plants, not only red and blue**





# **Thank you.**



# **Products**



# **OSLON® SSL COLOR New generation**









**Blue** 

Red



# **Appendix**



#### **Definitions**

**Radiometry:** deals with the detection and measurement of electromagnetic radiation across the total spectrum

**Photometry:** subfield of radiometry; radiometric power scaled by the spectral response of the human eye

**Photon Flux:** number of photons in a spectral range per unit time. When limited to the range 400-700 nm, it is termed Photosynthetic Photon Flux.

**Mol/mol/µmol:** In chemistry, a unit of measurement counting the number of atoms/molecules/electrons/etc. in a substance (for horticulture, photons) By definition, the number of photons in a mol is 6.022 x 10<sup>23</sup> (Avogadro's number)

**Photon:** Discrete bundle (quantum) of electromagnetic radiation (light). Can be considered to be a particle (although it displays properties of waves as well). The energy of a photon depends upon its wavelength. Conversely, if the energy & wavelength are known, the number of photons can be calculated

**Photosynthetically Active Radiation (PAR):** Radiation between 400 nm and 700 nm. Spectral region most useful to plants for photosynthesis

**Photosynthetic Photon Flux Density (PPFD):** Radiation between 400 nm and 700 nm. Radiation hitting a surface



#### **Definitions**

**Photosynthesis: A** process used by plants and other organisms to convert light energy into chemical energy that can be later released to fuel the organisms' activities. This chemical energy is stored in carbohydrate molecules, such as sugars, which are synthesized from carbon dioxide and water.

**Germination:** Germination is the process by which a plant grows from a seed. It is also known as sprouting of a seedling from a seed.

**Vegetative Growth:** Vegetative Growth is the period between germination and flowering. It is also known as vegetative phase of the plant development. During this phase the plants are performing photosysthesis and accumulating resources which will be used for the flowering and reproduction in the later stage.

**Photomorphogenesis:** Because light is the energy source for plant growth, plants have evolved highly sensitive mechanisms for perceiving light and using that information for regulating development changes to help maximize light utilization for photosynthesis. The process by which plant development is controlled by light is called photomorphogenesis. Typically, photomorphogenic responses are most obvious in germinating seedlings but light affects plant development in many ways throughout all stages of development.

**Flowering:** The transition to flowering is one of the major phase changes a plant makes during its life cycle. The transition must take place at a time that is favorable for fertilization and the formation of seeds. The right photoperiod is essential for the flowering.

**Etiolatio:** Abnormal shape of plants due to significantly accelerated length growths caused by insuficient illumination which can be used for photosynthesis.



References

0. Singh, Devesh, et al. "LEDs for Energy Efficient Greenhouse Lighting." arXiv preprint arXiv:1406.3016 (2014).

1. Mitchell CA, Both A, Bourget CM, Kuboto C, Lopez RG, Morrow RC & Runkle S. LEDs: The future of greenhouse lighting. Chronica Horticulture. 2012;55:6-12.

2. Morrow RC. LED lighting in horticulture. Hort Science. 2008;43:1947–1950.

3. Yeh N & Chung JP. High-brightness LEDs – energy efficient lighting sources and their potential in indoor plant cultivation. Renew Sust Energ Rev. 2009;13:2175–2180.

4. Tennessen DJ, Singsaas EL & Sharkey TD. Light-emitting diodes as a light source for photosynthesis research. Photosynth Res. 1994;39:85– 92.

5. Barta DJ, Tibbits TW, Bula RJ & Morrow, RC. Evaluation of light emitting diode characteristics for a space-based plant irradiation source. Adv Space Res. 1992;12:141–9.

6. Olle M & Virsile A. The effect of light-emitting diode lighting on greenhouse plant growth and quality. Agric Food Sci. 2013;22:223-234.

7. Li Q & Kubota C. Effects of supplemental light quality on growth and phytochemicals of baby leaf lettuce. Environ Exp Bot. 2009;67:59–64.

8. Lin KH, Huang MY, Huang WD, Hsu MH, Yang ZW & Yang CM. The effects of red, blue, and white light-emitting diodes on the growth, development, and edible quality of hidroponically grown lettuce (Lactuca sativa L. var. capitata). SciHortic-Amsterdam. 2013;150:86–91.

9. Massa GD, Kim HH, Wheeler RM & Mitchell CA. Plant productivity in response to LED lighting. Hort Science. 2008;43:1951–1956.

10. Vänninen I, Pinto DM, Nissinen AI, Johansen NS & Shipp L. In the light of new greenhouse technologies: Plant-mediates effects of artificial lighting on arthropods and tritrophic interactions. Ann Appl Biol. 2010;157:393–414.



11. Bourget CM. An introduction to light-emitting diodes. Hort Science. 2008;43:1944–1946.

12. Brumfield R. Dealing with rising energy costs. GPN. 2007;17:24-31.

13. Langton A, Plackett C & Kitchener H. Energy saving in poinsettia production. Horticultural Development Council Fact sheet. 2006;7:1-12.

14. Opdam JG, Schoonderbeek GG, Heller EB & Gelder A. Closed greenhouse: a starting point for sustainable entrepreneurship in horticulture. Acta Hort. 2005;691:517-524.

15. Ieperen VW & Trouwborst G. The Application of LEDs as Assimilation Light Source in Greenhouse Horticulture: a Simulation Study. Acta Hort. 2008;33:1407-1414.

16. Nanya K, Ishigami Y, Hikosaka S & Goto E. Effects of blue and red light on stem elongation and flowering of tomato seedlings. Acta Hort. 2012;956:261–266.

17. Keefe TJ. "The Nature of Light".Archived from the original on 2012-07-24. Retrieved 2007-11-05 Tower Hall Funabori, Tokyo, Japan.

18. Nishio JL. Why are higher plants green? Evolution of the higher plant photosynthetic pigment complement. Plant Cell Environ. 2000;23:539– 548.

19. Chen P. Chlorophyll and other photosentives. In: LED grow lights, absorption spectrum for plant photosensitive pigments. [http://www.ledgrowlightshq.co.uk/chlorophyll-plant-pigments/.](http://www.ledgrowlightshq.co.uk/chlorophyll-plant-pigments/) Accessed 12 March 2014.

20. Bula RJ, Morrow RC, Tibbits TW, Barta RW, Ignatius RW & Martin TS. Light emitting diodes as a radiation source for plants. Hort Science.1991;26:203–205.

21. Tanaka Y, Kimata K & Aiba H. A novel regulatory role of glucose transporter of Escherichia coli: membrane sequestration of a global repressor Mic. EMBO J. 2000;19:5344-5352.



.

22. Tripathy BC & Brown CS. Root-shoot interaction in the greening of wheat seedlings grown under red light. Plant Physiol. 1995;107:407–411.

23. Yanagi T & Okamoto K. Utilization of super-bright light emitting diodes as an artificial light source for plant growth. Acta Hort. 1997;418:223- 228.

24. Barreiro R, Guiamet JJ, Beltrano J & Montaldi ER. Regulation of the photosynthetic capacity of primary bean leaves by the red: far-red ratio and photosynthetic photon flux density of´incident light. Physiol. Plant. 1992;85:97–101

25. Sims DA & Pearcy RW. Response of leaf anatomy and photosynthetic capacity in Alocasiamacrorrhiza (Araceae) to a transfer from low to high light. Am J Bot. 1992;79:449–455.

26. Akoyunoglou G & Anni H. Blue light effect on chloroplast development in higher plants. In: Senger H. (ed.), Blue Light Effects in Biological Systems. Springer-Verlag, Berlin: 1984. pp. 397–406.

27. Saebo A, Krekling T & Appelgren M. Light quality affects photosynthesis and leaf anatomy of brich plantlets in vitro.Plant Cell Tiss Org. 1995;41:177–185.

28. Senger H. The effect of blue light on plants and microorganisms. Phytochem Photobiol. 1982;35:911–920.

29. Yorio NC, Goins GD, Kagie HR, Wheeler RM & Sager JC. Improving spinach, radish and lettuce growth under red light emitting didoes (LEDs) with blue light supplementation. Hort Science. 2001;36:380–383.

30. Stutte GW, Edney S & Skerritt T. Photoregulation of bioprotectant content of red leaf lettuc with light-emitting diodes. Hort Science. 2009;44:79–82.

31. Goins GD, Ruffe LM, Cranston NA, Yorio NC, Wheeler RM & Sager JC. Salad crop production under different wavelengths of red lightemitting diodes (LEDs). SAE Technical Paper, 31st International Conference on Environmental Systems, July 9–12, 2001, Orlando, Florida, USA: 2001. p. 1–9.



32. Li H, Tang C, Xu Z, Liu X & Han X. Effects of different light sources on the growth of nonheading chinese cabbage (Brassica campestris L.). J Agr Sci. 2012;4:262–273.

33. Mizuno T, Amaki W & Watanabe H. Effects of monochromatic light irradiation by LED on the growth and anthocyanin contents in laves of cabbage seedlings. Acta Horticulturae. 2011;907:179–184.

34. Brown C, Shuerger AC & Sager JC. Growth and photomorphogenesis of pepper plants under red light-emitting diodes with supplemental blue or far-red lighting. J Am SocHortic Sci. 1995;120:808–813.

35. Goins GD, Yorio NC, Sanwo MM & Brown CS. Photomorphogenesis, photosynthesis and seed yield of wheat plants grown under red lightemitting diodes (LEDs) with and without supplemental blue lighting. J Exp Bot. 1997;48:1407–1413.

36. Lefsrud MG, Kopsell DA & Sams CE. Irradiance from distinct wavelength light-emitting diodes affect secondary metabolites in kale. Hort Science. 2008;43:2243–2244.

37. Johkan M, Shoji K, Goto F, Hahida S & Yoshihara T. Effect of green light wavelength and intensity on photomorphogenesis and photosynthesis in Lactuca sativa. Environ Exp Bot. 2012;75:128–133.

38. Tarakanov I, Yakovleva O, Konovalova I, Paliutina G & Anisimov A. Light-emitting diodes: on the way to combinatorial lighting technologies for basic research and crop production. ActaHorticulturae. 2012;956:171–178.

39. Lu N, Maruo T, Johkan M, Hohjo M, Tsukakoshi S, Ito Y, Ichimura T & Shinohara Y. Effects of supplemental lighting with light-emitting diodes (LEDs) on tomato yield and quality of single-truss tomato plants grown at high planting density. Environ Control Biol. 2012;50:63– 74.

40. Samuolienė G, Urbonavičiūtė A, Duchovskis P, Bliznikas Z, Vitta P & Žukauskas A. Decrease in nitrate concentration in leafy vegetables under a solid-state illuminator. Hort Science. 2009;44:1857–1860.



41. Bliznikas Z, Žukauskas A, Samuolienė G, Viršilė A, Brazaitytė A, Jankauskienė J, Duchovskis P & Novičkovas A. Effect of supplementary pre-harvest LED lighting on the antioxidant and nutritional properties of green vegetables. Acta Hort. 2012;939:85–91.

42. Samuolienė G, Sirtautas R, Brazaitytė A, Viršilė A & Duchovskis P. Supplementary red-LED lighting and the changes in phytochemical content of two baby leaf lettuce varieties during three seasons. J Food Agric Environ. 2012a;10:701 – 706.

43. Samuolienė G, Brazaitytė A, Sirtautas R, Novičkovas A & Duchovskis P. Supplementary red-LED lighting affects phytochemicals and nitrate of baby leaf lettuce. J Food Agric Environ. 2011;9:271–274.

44. Žukauskas A, Bliznikas Z, Breivė K, Novičkovas A, Samuolienė G, Urbonavičiūtė A, Brazaitytė A, Jankauskienė J & Duchovskis P. Effect of supplementary pre-harvest LED lighting on the antioxidant properties of lettuce cultivars. Acta Hort. 2011;907:87–90.

45. Ménard C, Dorais M, Hovi T & Gosselin A. Developmental and physiological responses of tomato and cucumber to additional blue light. Acta Hort. 2006;711:291–296.

46. Novičkovas A, Brazaitytė A, Duchovskis P, Jankauskienė J, Samuolienė G, Viršilė A, Sirtautas R, Bliznikas Z & Žukauskas A. Solid-state lamps (LEDs) for the short-wavelength supplementary lighting in greenhouses: experimental results with cucumber. Acta Hort. 2012;927:723-730.

47. Samuolienė G, Brazaitytė A, Duchovskis P, Viršilė A, Jankauskienė J, Sirtautas R, Novičkovas A, Sakalauskienė S & Sakalauskaitė,J. Cultivation of vegetable transplants using solid-state lamps for the short-wavelength supplementary lighting in greenhouses. Acta Hort. 2012c ;952:885–892.

48. Folta KM. Green light stimulates early stem elongation, antagonizing light-mediated growth inhibition. Plant Physiol. 2004;135:1407–1416.

49. Kim HH, Goins GD, Wheeler RM & Sager JC. Green- light supplementation for enhanced lettuce growth under red and blue light-emitting diodes. Hort Science. 2004;39:1617–1622.

50. Simpson GG & Dean C. Arabidopsis, the Rosetta stone of flowering time? Science. 2002; 296:285–289.



51.Yanovsky MJ & Kay SA. Molecular basis of seasonal time measurement in Arabidopsis. Nature. 2002;419:308–312.

52. Downs RJ & Thomas JF. Phytochrome regulation of flowering in the long-day plant, Hyoscyamusniger. Plant Physiol. 1982;70:898–900.

53. Evans LT. Inflorescence initiation in Loliumtemu lentum L. XIV. The role of phytochrome in long day induction. Austral. J. Plant Physiol. 1976;3:207–217.

54. Shinomura T, Uchida K & Furuya M. Elementary processes of photoperception by phytochrome A for high-irradiance response of hypocotyl elongation in Arabidopsis. Plant Physiol. 2000;122:147–156.

55. Smith H. Light quality, photoperception, and plant strategy. Annu Rev Plant Physiol. 1982;33:481–518.

56. Runkle ES & Heins DR. Specific functions of red, far-red and blue lights in flowering and stem extension of long-day plants. J Amer Soc. Hort Sci. 2001;126:275–282.

57. Meng Q & Runkle ES. Control flowering with LEDs. Lighting Research.Growers Talk 62., http://www.ballpublishing.com/GrowerTalks/ViewArticle.aspx?articleid=20604 Accessed 15 Feb 2014.

58. Gomez C, Morrow RC, Bourget CM, Massa GD & Mitchell CA. Comparison of intracanopy light-emitting diode towers and overhead highpressure sodium lamps for supplemental lighting of greenhouse-grown tomatoes. Hort Technology. 2013;23:93–98.

59. Voss J. Market special: greenhouse farming in Germany. The ministry of Economics Affairs, Agriculture and Innovation, NL, EVD International. 2011. http://duitsland.nlambassade.org/binaries/content/assets/postenweb/d/duitsland/ambassadeberlijn/zaken-doen/20110507 marktverkenning-greenhouse-farming-germany.pdf Accessed 16 Feb 2014

60. Kacira, M. Greenhouse Production in US: Status, Challenges, and Opportunities. Presented a CIGR 2011 conference on Sustainable Bioproduction WEF 2011, September 19-23, 2011. 61. Nelson AJ & Bugbee B. 2013. Supplemental greenhouse lighting: Return on investments for LED and HPS fixtures. http://cpl.usu.edu/files/publications/factsheet/pub\_\_4338884.pdf



# **Thank you.**

